Clinical, biochemical, and genetic analysis of a Korean neonate with hereditary tyrosinemia type 1

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Abstract

Background: Hereditary tyrosinemia type 1 (HT1; MIM 276700) is caused by mutations in the fumarylacetate hydrolase (FAH) gene, and is the most severe disorder associated with the tyrosine catabolic pathway. HT1 is a very rare disorder and no genetically confirmed case of HT1 in Korea has yet been reported. In this study, we present a Korean neonate with clinical and biochemical features of HT1.

Methods: A female neonate was admitted to our hospital for further work-up of an abnormal newborn screening test. We analyzed amino acids and organic acids in the patient’s blood and urine. To confirm the presence of the genetic abnormality, all the coding exons of the FAH gene and the flanking introns were amplified by polymerase chain reaction (PCR).

Results: The patient’s newborn screening test revealed increased concentrations of methionine and tyrosine. Subsequent urine organic acid analysis showed increased urinary excretion of 4-hydroxyphenyllactate, 4-hydroxyphenylpyruvate, succinate, and succinylacetone. Gap-PCR and sequence analysis of the FAH gene revealed a homozygous large deletion mutation encompassing exons 12–14. The patient’s parents were not consanguineous but were heterozygous carriers of the same mutation.

Conclusions: The patient had a novel, large deletion mutation of FAH and is the first report of genetically confirmed HT1 in Korea.

Keywords: fumarylacetoacetate hydrolase (FAH); Korean; large deletion; novel mutation; tyrosinemia.

Introduction

Hereditary tyrosinemia type I (HT1, MIM 276700), also referred to as hepato-renal tyrosinemia, is the most severe disorder linked to the tyrosine catabolic pathway (1). HT1 is caused by decreased activity of fumarylacetoacetate hydrolase (FAH; EC 3.7.1.2), which is the last enzyme that hydrolyzes fumarylacetoacetate to fumarate and acetocacetate in the tyrosine catabolic pathway (2, 3). FAH deficiency causes severe progressive liver disease in infancy, renal tubular defects with hypophosphatemic rickets, and neurologic crises (2). If not treated, patients usually die of liver failure within the first year of life.

The worldwide prevalence of HT1 is known to be very low (1:100,000–1:120,000 births); its prevalence in Koreans is not known (4, 5). The human FAH gene maps to the long arm of chromosome 15 in the region q23–q25 and contains 14 exons that encode FAH mRNA with a length of at least 1477 nucleotides (6, 7). There are variable levels of FAH enzyme activity in liver tissue from patients with HT1 (8). The relationship between genotype and phenotype is unclear, because different clinical presentations of HT1 have been reported in patients with identical genotypes (9). Genotype heterogeneity is not sufficient for explaining the clinical heterogeneity, and other factors may modify the phenotype in HT1 (9).

In this report, we present the first biochemically and genetically confirmed Korean patient with HT1. Interestingly, the patient was homozygous for a novel large deletion mutation of the FAH gene.

Materials and methods

Clinical and biochemical analysis

A female neonate was admitted to our hospital for further evaluation of an abnormal newborn screening test performed 2 days after birth at a local hospital. At 1 month of age, the patient underwent various biochemical tests, including total protein, aspartate aminotransferase, alanine aminotransferase, alkaline phosphatase, ammonia, albumin, bilirubin and lactate. In addition, tests in the newborn screening test were confirmed by liquid chromatography-
tandem mass spectrometry (LC-MS/MS), and amino acids and organic acids in the blood and urine were analyzed. Abdominal ultrasonography was performed to visualize abdominal anatomical structures.

**Genetic analysis**

The study was approved by the Ethical Committee of our institution. Blood samples were collected from the patient and parents after obtaining informed parental consent. Genomic DNA was isolated from peripheral blood leukocytes using the Wizard Genomic DNA Purification Kit (Promega, Madison, WI, USA). All of the coding exons of the FAH gene and flanking introns were amplified using polymerase chain reaction (PCR) with primers designed by the authors (sequences available upon request) and a thermal cycler (Model 9700; Applied Biosystems, Foster City, CA, USA). Direct sequencing was performed with the ABI Prism 3100 Genetic Analyzer (Applied Biosystems) using the BigDye Terminator Cycle Sequencing-Ready Reaction Kit (Applied Biosystems). All novel mutations were confirmed by testing 100 control chromosomes.

**Gap-PCR**

Gap-PCR using the AccuPower TLA PCR premix (Bioneer, Daejeon, Korea) was performed since the patient had a large deletion in the FAH gene. Using chromosomal walking with different pairs of primers, the smallest PCR amplicon (expected size: 18,453 bp) was detected using the primer pair: forward (5’-acctgtgctctttgcag-3’) and reverse (3’-gaaggtatccaaaccaagtga-5’). The primer sequences were based on the intron sequences of the FAH gene obtained from the Ensembl Genome Browser (http://www.ensembl.org).

**Results**

**Clinical and biochemical findings**

The patient was born at 40 weeks’ gestation by vaginal delivery and weighed 3.48 kg. Physical examination revealed an asymmetric face and hyperflexible wrist joints. A repeat newborn screening test at 9 days of life showed increased concentrations of phenylalanine, methionine and tyrosine: 155 μM phenylalanine (cut-off, ≤125 μM), 199 μM methionine (cut-off, ≤67 μM) and 481 μM tyrosine (cut-off, ≤305 μM).

Liver function tests performed at 1 month of age were also abnormal. Aspartate aminotransferase was 0.94 [reference range (RR) <0.53] μkat/L, alkaline phosphatase was 1650 (RR, 42–98) U/L, ammonia was 89.3 (RR, 17.9–46.4) μmol/L, and lactate was 6.5 (RR, 0.5–2.2) mmol/L. Urine organic acid analysis at 1 month of age showed increased urinary excretion of 4-hydroxyphenyllactate, 4-hydroxyphenylpyruvate, succinate, and succinylacetone [4-hydroxyphenyl-lactate: 893 mmol/mol creatinine (cut-off, <3), 4-hydroxyphenylpyruvate: 563 mmol/mol creatinine (cut-off, <1), succinate: 370 mmol/mol creatinine (cut-off, <79), and succinylacetone: 28 mmol/mol creatinine (cut-off, non-detected)]. Abdominal ultrasound revealed ascites and coarse parenchymal changes in the liver.

The patient was diagnosed with HT1 based on clinical and biochemical tests at 47 days after birth. Treatment with 2-(2-nitro-4-trifluoromethylbenzoyl)-1,3-cyclohexanedione (NTBC) and dietary tyrosine restriction was initiated after confirmation of tyrosinemia type I. Following 1 week of treatment, the patient’s increased urinary succinylacetone declined to non-detectable concentrations and remained <1 mmol/mol creatinine during follow-up. Urinary concentrations of methionine and tyrosine reverted to normal values of 5 μM and 11 μM, respectively, by 2 months of age. The patient’s disease was well controlled, and she was seen as an outpatient.

At 4 months after birth, the patient developed acute symptoms, including uncontrolled fever and dyspnea suggestive of sepsis secondary to aspiration pneumonia. This developed independently of the patient’s tyrosinemia. Although prompt management and close observation was performed, septic complications including metabolic acidosis, ascites, and disseminated intravascular coagulation developed and the patient expired following cardiac arrest.

**Genetic analysis**

PCR of each FAH exon with intron primer failed to amplify exons 12, 13, and 14, suggesting that the patient was homozygous for a large deletion. Using different primer pairs that covered the region from intron 11 of the FAH gene to the FAH-ARNT2 intergenic region, gap-PCR was performed to identify the breakpoint of the large deletion of exons 12, 13 and 14 (Figure 1). Sequence analysis of the PCR amplicon revealed an 18,036 bp deleted region that spanned from nucleotide 1130 of intron 11 of the FAH gene to nucleotide 10,539 of the FAH-ARNT2 intergenic region. According to the numbering position of the human FAH reference sequence, the mutation could have resulted in an expected amplicon of 18.5 kb, which was not amplified from a control sample. A shorter 417 bp product was detected in the patient, who has a homozygous large deletion. The patient’s parents were carriers of this large deletion.

![Figure 1](image-url)
be described as c.960+1130_*1260+10539del18036 (reference sequence from NC_000015.8 and NM_000137.1) (Figure 2). The patient’s parents were carriers of this large deletion, and the patient inherited one mutant allele from each parent. Screening of 50 normal controls revealed no mutant allele in 100 chromosomes.

**Discussion**

Currently, most laboratories perform newborn screening tests using dried blood spots analyzed with LC-MS/MS. However, this is a limited screening method for HT1 because tyrosine is increased in cases of benign transient tyrosinemia. In addition, the increased concentration of tyrosine in patients with HT1 may overlap the normal range in control populations (4). Therefore, we determined the concentration of succinylacetone, a specific marker for HT1, in dried blood spots using LC-MS/MS (10). The concentrations of methionine and tyrosine in this case were not dramatically increased, but we diagnosed HT1 with the results from the analysis of urine organic acids.

Although there have been a few reports of Korean patients with tyrosinemia type 1 (11–13), all cases were diagnosed by clinical and biochemical findings without investigation of the molecular characteristics. This is the first case in Korea of HT1 that was confirmed with genetic analysis in addition to the biochemical abnormalities. In addition, the large deletion identified in this study is a novel mutation. The most common FAH gene mutations in patients with HT1 differ with respect to ethnicity. For example, the specific mutation IVS12+5G>A is prevalent in French Canadian and Scandinavian, excluding Finnish, populations (14, 15). In addition, five mutations IVS6-1G>T, c.1009G>A, c.192G>T (a splicing error), p.D233V, and p.W262X are common in Central and Western Europe, Scandinavia, Pakistan, Turkey, and Finland, respectively (15–18). These mutations are thought to have spread by the founder effect.

In the FAH gene, missense and nonsense point mutations, and splicing errors are common and account for 90% of the total number of mutations reported in the Human Gene Mutation Database (HGMD) at the Institute of Medical Genetics in Cardiff (http://www.hgmd.cf.ac.uk/ac/gene.php?gene=FAH). Large deletions of the FAH gene have rarely been reported. Only one mutant allele with a large deletion, E6/I6del26 (c.548_553+20del), has been reported previously (17). In the present study, we identified a novel mutation, c.960+1130_*1260+10539del18036, resulting in the loss of 100 residues (amino acids 321–420) from the protein. It was also interesting that the patient’s parents were heterozygous carriers of the same allele with a large deletion of the FAH gene. The presence of a homozygous deletion in a non-consanguineous pedigree is very unusual.

![Chromosome 15q25](image)

**Figure 2** Breakpoint analysis of the deletion of exons 12–14 of the FAH gene.

Sequence analysis shows a deletion of 18,036 bp that spans from nucleotide 1130 of intron 11 of the FAH gene to nucleotide 10,539 of the FAH-ARNT2 intergenic region, designated as c.960+1130_*1260+10539del18036 (reference sequence from NC_000015.8 and NM_000137.1).
Although the patient temporarily improved after administration of NTBC, she died at 4 months of age due to aspiration pneumonia and sepsis. NTBC is a potent inhibitor of 4-hydroxyphenylpyruvate dioxygenase and has improved the outcomes of most patients with hereditary tyrosinemia (19). The patient was diagnosed clinically with an acute form of HT1, given her presentation of hepatic signs and symptoms within the first 6 months of life (20). This clinical presentation may be due to the large FAH deletion, containing exons 12–14, which lead to the synthesis of structurally unstable, incomplete proteins, thereby inducing attenuation of FAH activity.

In summary, we diagnosed a Korean patient with HT1 using biochemical and molecular analysis, revealing a novel large-deletion FAH mutation. The gap-PCR method developed in this study may be a useful tool for the identification of Korean HT1 patients with deletions of exons 12–14.

References